

Non-invasive assessment of liver fibrosis in chronic hepatitis C

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Abstract Quantification of hepatic fibrosis is of critical importance in chronic hepatitis C not only for prognosis, but also for antiviral treatment indication. Two end points are clinically relevant: detection of significant fibrosis (indication for antiviral treatment) and detection of cirrhosis (screening for esophageal varices and hepatocellular carcinoma). Until recently, liver biopsy was considered the reference method for the evaluation of liver fibrosis. Limitations of liver biopsy (invasiveness, sampling error, and inter-observer variability) have led to the development of non-invasive methods. Currently available methods rely on two different approaches: a “biological” approach based on the dosage of serum fibrosis biomarkers; and a “physical” approach based on the measurement of liver stiffness, using transient elastography (TE). This review is aimed at discussing the advantages and limits of non-invasive methods and liver biopsy and the perspectives for their rational use in clinical practice in the management of patients with chronic hepatitis C.

Keywords Chronic hepatitis C · Liver fibrosis · Non-invasive · Transient elastography · FibroScan · Serum biomarkers · Liver biopsy

Introduction

In patients with chronic hepatitis C, precise staging of liver fibrosis is important not only for estimation of prognosis, but also for indication of antiviral therapy. Two end points are clinically relevant: detection of significant fibrosis, which is an indication for antiviral treatment, and detection of cirrhosis, which is an indication for specific monitoring of complications related to portal hypertension and to the increased risk of developing hepatocellular carcinoma [1].

Liver biopsy has traditionally been considered the reference method for the evaluation of liver fibrosis in chronic hepatitis C [2]. However, liver biopsy is an invasive procedure, associated with pain in around 30% of cases [3–5], and potentially life-threatening complications (hemorrhage in 0.3% of cases and mortality in 0.01%) [6]. The accuracy of liver biopsy to assess fibrosis has also been questioned in relation to sampling errors and intra- and inter-observer variability that may lead to over- or under-staging [7–10]. Finally, liver biopsy does not allow dynamic evaluation of liver fibrosis over time.

These limitations have led to the development of non-invasive methods [11]. Currently available methods rely on two different, but complementary, approaches: [1] a “biological” approach based on the dosage of serum biomarkers of fibrosis; and [2] a “physical” approach based on the measurement of liver stiffness, using transient elastography (TE) [12]. Although the large number of publications over the past decade confirms the growing interest regarding these new non-invasive methods, there is a need for clarification and guidance on their use and interpretation. This review is aimed at discussing the advantages and limits of these methods, and the perspectives for their rational use in clinical practice in the management of patients with chronic hepatitis C.

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Biological approach: serum fibrosis biomarkers

Individual surrogate serum markers of liver fibrosis

A variety of “direct” serum markers of fibrosis, reflecting either the deposition or the removal of extracellular matrix in the liver have been evaluated for their ability to assess liver fibrosis. These include: glycoproteins, such as serum hyaluronate [13–17], laminin [18, 19], and YKL-40 [20]; the collagens family, such as procollagen III N-peptide (PIINP) [13] and type IV collagen [21]; collagenases and their inhibitors, such as matrix metalloproteases (MMP) [22] and tissue inhibitory metalloprotease-1 (TIMP-1) [23].

“Indirect” serum markers including simple routine blood tests, such as prothrombin index [19], platelet count [24], and AST/ALT ratio [25–27] have also been proposed.

The performances of “direct” and “indirect” serum markers have been evaluated in chronic hepatitis C (reviewed in [12, 28–30]), with hyaluronate being the most extensively studied. Apart from hyaluronate [14, 15, 31], “direct” and “indirect” markers, when used individually, are useful for the diagnosis or the exclusion of cirrhosis, but have limited accuracy for the diagnosis of significant fibrosis [32].

Scores combining indirect and direct markers of liver fibrosis

The limitations of individual markers to assess liver fibrosis have led to the development of more sophisticated scores combining the results of panels of markers that substantially improved diagnostic accuracy for which the FibroTest has been the pioneer [33]. Several other scores have been developed since then [34–48] (Table 1). The diagnostic performances for significant fibrosis and cirrhosis of proposed scores are summarized in Tables 2 and 3, respectively.

Five scores are protected by patents and are currently commercially available: the FibroTest® in Europe (Biopredictive, Paris, France) licensed under the name of Fibrosure® in the USA (LabCorp, Burlington, NC, USA), the Fibrometers® (BioLiveScale, Angers, France), the FibroSpectII® (Prometheus Laboratory Inc. San Diego, CA, USA), the ELF® (Enhanced Liver Fibrosis Test, iQur Ltd, Southampton, UK), and the Hepascore® (PathWest, University of Western Australia, Australia).

To date, FibroTest® and aspartate to platelet ratio index (APRI) have been the most extensively studied. In a meta-analysis [49], which pooled 6,378 subjects (with analysis of individual data in 3,282) with both FibroTest and biopsy (3,501 HCV), the mean standardized area under the ROC curve (AUROC) for diagnosing significant fibrosis was

0.84 (95% CI, 0.83–0.86). In another meta-analysis [50], which pooled 4,266 HCV patients from 22 studies, the mean AUROCs of APRI for diagnosing significant fibrosis and cirrhosis were 0.76 (0.74–0.79) and 0.82 (0.79–0.86), respectively.

When compared and validated externally in patients with hepatitis C [51–56], the different patented scores have similar performances for the diagnosis of significant fibrosis. In the largest study to date ($n = 1,307$) [56], comparing prospectively several patented and non-patented scores (FibroTest®, Fibrometre®, Hepacore® and APRI), the AUROCs ranged from 0.72 to 0.78 for significant fibrosis and from 0.77 to 0.86 for cirrhosis. Although non-patented scores, such as the Forns index, FIB-4, and APRI may have slightly lower performance, they are cost-free, easy to calculate, and available almost everywhere.

Limitations

One of the main limitations to the clinical use of “direct” markers of liver fibrosis is that they are not routinely available in most hospital settings. Another limitation of these markers is that none is liver-specific and they may be influenced by changes in their clearance and excretion. For instance, increased levels of hyaluronate occur in the postprandial state [57] or in aged patients with chronic inflammatory processes such as rheumatoid arthritis. Also, the reproducibility of measurement of some parameters included in “indirect” serum markers, such as AST levels or platelet count, is questionable [58]. Conversely, the inter-laboratory reproducibility of scores, such as FibroTest® and Fibrometers® has been shown to be satisfactory for use in clinical practice [59–61]. However, when using FibroTest®, the interpretation should be done critically, taking into account each of the five components individually to avoid false-positive results related to hemolysis (decrease in haptoglobin) and Gilbert syndrome (increase in bilirubin), or false-negative results related to inflammation [62].

Physical approach: transient elastography

Principle

TE (FibroScan®, Echosens, Paris, France) has been proposed for the measure of liver stiffness [63]. Briefly, an ultrasound transducer probe is mounted on the axis of a vibrator. Vibrations of mild amplitude and low frequency are transmitted by the transducer, inducing an elastic shear wave that propagates through the underlying tissues. Pulse-echo ultrasound acquisitions are used to follow the propagation of the shear wave and to measure

Table 1 Proposed serum scores for non-invasive evaluation of liver fibrosis in chronic hepatitis C

- Fibrotest® (α -2-macroglobulin, γ GT, apolipoprotein A1, haptoglobin, total bilirubin, age, and gender)
- Forns Index (age, platelet count, cholesterol, and γ GT)
- AST to platelet ratio (APRI) (AST and platelet count)
- FibroSpect® (α -2-macroglobulin, hyaluronate, and TIMP-1)
- MP3 (MMP-3 and TIMP-1)
- Enhanced Liver Fibrosis score® (ELF) (age, hyaluronate, MMP-3, and TIMP-1)
- Fibrosis Probability Index (FPI) (age, past alcohol intake, AST, cholesterol, and HOMA-IR)
- Lok Index (platelet count, AST/ALT ratio, and INR)
- Goteborg University Cirrhosis Index (GUCI) (AST, INR, and platelet count)
- Hepascore® (bilirubin, γ GT, hyaluronate, α -2-macroglobulin, age, and gender)
- Fibrometers® (platelet count, prothrombin index, AST, α -2-macroglobulin, hyaluronate, urea, and age)
- Virahep-C model (AST, platelet count, alkaline phosphatase, and age)
- Fibroindex (platelet count, AST, and γ -globulin)
- FIB-4 (Platelet count, ALT, and AST)
- HALT-C model (hyaluronic acid, TIMP-1, and platelet count)

Table 2 Diagnostic performance of non-invasive serum scores of liver fibrosis for significant fibrosis in chronic hepatitis C

Scores	Year	Patients (n)	Significant fibrosis (%)	Cutoffs	AUROC	Se	Sp	+LR	-LR
FibroTest® [33]	2001	339	80	>0.48	0.87	75	85	5	0.3
Forns Index [34]	2002	476	26	<4.2 >6.9	0.76–0.86	30–94	51–95	1.9–6.0	0.1–0.7
APRI [35]	2003	270	50	$\leq 0.5 >1.5$	0.69–0.88	41–91	47–95	1.7–8.2	0.2–0.6
FibroSpect® [36]	2004	696	52	>0.36	0.83	77	73	2.9	0.3
MP3 [37]	2004	194	45	<0.3 >0.4	0.82	35–65	85–96	4.3–8.8	0.4–0.7
ELF® [38]	2004	1,021 ^a	40	NA	0.78	90	30	1.3	0.3
FPI [39]	2005	302	48	<0.2 ≥ 0.8	0.77	42–85	48–98	1.6–21.0	0.3–0.6
Hepascore® [40]	2005	211	57	≥ 0.5	0.82	63	89	5.7	0.4
Fibrometer® [41]	2005	598 ^b	56	NA	0.89	80	84	5	0.2
ViraHep-C [42]	2006	859	37	$\leq 0.22 >0.55$	0.83	51–90	54–90	2.0–5.1	0.2–0.5
Fibroindex [43]	2007	360	50	$\leq 1.25 \geq 2.25$	0.83	30–40	97–97	10.0–13.3	0.6–0.7

AUROC Area under ROC curve, Se sensitivity, Sp specificity, +LR positive likelihood ratio, -LR negative likelihood ratio, NA not available

^a 496 HCV patients; ^b 383 with viral hepatitis in the exploratory population and 120 HCV in the validation population

Table 3 Diagnostic performance of serum non-invasive scores of liver fibrosis for cirrhosis in chronic hepatitis C

Scores	Year	Patients (n)	Cirrhosis (%)	Cut-offs	AUROC	Se (%)	Sp (%)	+LR	-LR
FibroTest® [56]	2001	1,197	14	>0.74	0.82	63	84	4	0.4
APRI [35]	2003	476	17	$<1.0 \geq 2.0$	0.94	57–89	75–93	3.6–8.1	0.1–0.5
Glycocirrhotos [44]	2004	106 ^a	45 ^b	NA	0.87	79	86	5.6	0.2
ELF® [38]	2004	1,021 ^c	NA	NA	0.89	91	69	2.9	0.4
Lok Index [45]	2005	1,141	38	$<0.2 \geq 0.5$	0.81	40–98	53–99	2.1–40.0	0.04–0.6
Hepascore® [40]	2005	211	16	>0.84	0.89	71	89	6.5	0.3
GUCI [46]	2005	179	12	>1.0	0.85	80	78	3.6	0.3
FIB-4 [47] ^d	2007	847	17\$	$<1.45 >3.25$	0.85	38–74	81–98	3.9–19.0	0.3–0.6
HALT-C model [48]	2008	512	38	$<0.2 >0.5$	0.81	47–88	45–92	1.6–5.9	0.3–0.6

AUROC Area under ROC curve, Se sensitivity, Sp specificity, +LR positive likelihood ratio, -LR negative likelihood ratio, NA not available

^a 71 HCV patients; ^b 24% with decompensated cirrhosis; ^c 496 HCV patients; ^d F3F4 patients

its velocity, which is directly related to tissue stiffness of the elastic modulus: the stiffer the tissue, the faster the shear wave propagates. TE measures liver stiffness in a volume that approximates a cylinder of 1-cm wide and 4-cm long, between 25 and 65 mm below the skin surface. This volume is at least 100 times bigger than a biopsy sample and is therefore far more representative of the hepatic parenchyma. TE is painless, rapid (less than 5 min), and easy to perform at the bedside or in the outpatient clinic. The results are immediately available and expressed in kilopascals (kPa), corresponding to the median value of ten validated measurements and range from 2.5 to 75 kPa [64], with normal values around 5.5 kPa [65].

Reproducibility

Reproducibility is an important prerequisite for a widespread application of TE in clinical practice. TE reproducibility has been shown to be excellent for both inter-observer and intra-observer agreement, with intra-class correlation coefficients (ICC) of 0.98 [66, 67]. However, inter-observer agreement was significantly reduced in patients with lower degrees of hepatic fibrosis, with hepatic steatosis, and with increased body mass index [66], as well as for liver stiffness values <9 kPa [67].

Diagnostic performance

The two index studies suggesting the interest of TE in the assessment of liver fibrosis were conducted in patients with chronic hepatitis C [68, 69]. Many other groups have confirmed these results [56, 70–72] with a strong correlation of liver stiffness values with Metavir fibrosis stages and AUROCs ranging from 0.75 to 0.91 for the diagnosis of significant fibrosis and from 0.90 to 0.98 for cirrhosis. Cutoff values with optimal diagnostic accuracy were defined for each stage of fibrosis (Tables 4, 5). It should be stressed, however, that despite high AUROC values, a substantial overlap of liver stiffness values was observed

between adjacent stages of hepatic fibrosis, particularly for lower fibrosis stages.

Three meta-analyses recently addressed the issue of diagnostic performance of TE [73–75]. In the most comprehensive meta-analysis, based on 50 studies (15 full papers and 35 abstracts) [75], the mean AUROC was 0.84 (95% confidence interval (CI), 0.82–0.86) for the diagnosis of significant fibrosis with a suggested optimal cutoff of 7.6 kPa, and 0.94 (95% CI, 0.93–0.95) for the diagnosis of cirrhosis with a suggested optimal cutoff of 13.0 kPa. Meta-analyses with individual data are awaited.

TE appears to be a reliable method for the diagnosis of cirrhosis, better at excluding than at predicting cirrhosis. For instance, in a population of 1,007 patients with different chronic liver diseases, a cutoff value of 14.6 kPa yielded positive and negative predictive values of 74 and 96%, respectively [76].

Monitoring of disease progression

Another promising application of TE is for monitoring the progression of liver fibrosis. The clinical significance of the wide range of liver stiffness values observed in patients with cirrhosis (13–75 kPa) remains uncertain. Preliminary results suggested that liver stiffness values in cirrhotic patients increased as the liver disease progressed [77]. For instance, cutoff values of 27.5, 37.5, 49.1, 53.7, and 62.7 kPa, respectively, had >90% negative predictive value for the presence of stage 2/3 esophageal varices, Child–Pugh score B or C, and a past history of ascites, HCC, and esophageal bleeding. Although retrospective and conducted in a single centre, this study provided the first “proof of concept” that liver stiffness values may have prognostic value in a context of cirrhosis. In addition, several studies [78, 79] have shown a correlation between liver stiffness values and portal hypertension diagnosed by means of hepatic venous pressure gradient (HVPG) measurement, the gold standard for the diagnosis and staging of portal hypertension and a reliable predictor of clinical decompensation [80]. A correlation between liver stiffness

Table 4 Diagnostic performance of transient elastography for significant fibrosis in chronic hepatitis C

Authors	Patients (n)	Significant fibrosis (%)	Cutoffs (kPa)	AUROC	Se (%)	Sp (%)	+LR	-LR
Degos et al. [56]	913	62	5.2	0.75	90	32	1.3	0.3
Sporea et al. [72]	191	84	6.8	0.77	60	93	8.6	0.4
Castera et al. [69]	183	74	7.1	0.83	67	89	6.1	0.4
Lupsor et al. [71]	324	65	7.4	0.86	76	84	4.8	0.3
Arena et al. [70]	150	56	7.8	0.91	83	82	4.6	0.2
Ziol et al. [68]	251	65	8.6	0.79	56	91	6.6	0.5

AUROC Area under ROC curve, Se sensitivity, Sp specificity, +LR positive likelihood ratio, -LR negative likelihood ratio

Table 5 Diagnostic performance of transient elastography for cirrhosis in chronic hepatitis C

Authors	Patients (<i>n</i>)	Cirrhosis (%)	Cutoffs (kPa)	AUROC	Se (%)	Sp (%)	+LR	-LR
Lupsor et al. [71]	324	21	11.9	0.94	87	91	9.7	0.1
Castera et al. [69]	183	25	12.5	0.95	87	91	9.7	0.1
Castera et al. [83]	298	23	12.5	0.96	83	95	16.6	0.2
Degos et al. [56]	913	14	12.9	0.9	72	89	6.8	0.3
Ziol et al. [78]	251	19	14.6	0.97	86	96	23.1	0.1
Arena et al. [70]	150	19	14.8	0.98	94	92	11.3	0.1

AUROC Area under ROC curve, Se sensitivity, Sp specificity, +LR positive likelihood ratio, -LR negative likelihood ratio

values and the presence of esophageal varices has also been reported [79, 81–84]. However, liver stiffness measurement (LSM) cannot yet confidently predict the presence of esophageal varices in clinical practice and thus avoid the need for upper GI endoscopic screening of cirrhotic patients [85].

TE could be also valuable for assessing the severity of recurrent hepatitis C after liver transplantation, reducing the need for follow-up liver biopsies [78, 86–90]. It has been recently suggested that TE may perform better for significant fibrosis than serum (direct and indirect) biomarkers [91].

Finally, TE could be useful for evaluating fibrosis regression in patients with chronic hepatitis C achieving sustained viral eradication [92–94] or for monitoring fibrosis progression in untreated patients.

Long-term prospective follow-up studies are now awaited to see whether liver stiffness values can predict the occurrence of clinical events in patients with compensated cirrhosis. Interestingly, a recent Japanese prospective study has shown in a large cohort of patients with chronic hepatitis C a correlation between liver stiffness values and the risk of hepatocellular carcinoma [95]. Although these findings need to be confirmed in other settings (Caucasian patients or HBV-infected patients) and with longer follow-up, they suggest that TE could be used as a rapid screening tool to allocate cirrhotic patients to specific risk categories [96].

Limitations

The interpretation of TE results should be always in the hands of an expert clinician and should be made having at disposition information regarding patient demographics, disease etiology, and essential laboratory parameters, as well as carefully following the manufacturer's recommendations (number of valid shots ≥ 10 ; success rate (the ratio of valid shots to the total number of shots) $\geq 60\%$; and interquartile range (IQR, reflecting the variability of measurements) less than 30% of the median LSM value (IQR/LSM $\leq 30\%$) [64]). Indeed, in a recent prospective 5-year

study based on more than 13,000 examinations, LSM failure (no value after ten measurements or more) has been shown to occur in 3.1% of cases and LSM results to be uninterpretable (not meeting manufacturer's recommendations) in an additional 15.8% [97]. Limited operator experience, obesity, and more generally features of the metabolic syndrome were the main determinants of LSM failure or unreliable results. This challenges previous claims [67, 98] that LSM requires no learning curve and that a novice can consistently obtain reliable results after a short training period. Operator experience influenced not only the success rate, as previously reported [67, 98], but also the IQR/LSM ratio, which was recently shown to be critical for LSM accuracy [99]. These findings emphasize the need for rigorous scientific evaluation of all novel procedures, such as TE before their widespread uptake. Further technological refinements are required for specific populations, such as those with the metabolic syndrome.

Finally, as the liver is an organ wrapped in a distensible but non-elastic envelope (Glisson's capsula), additional space-occupying tissue abnormalities, such as edema and inflammation, cholestasis and congestion may interfere with LSM, independently of fibrosis. Indeed, as previously mentioned, the extent of necro-inflammatory activity has been shown to influence TE measurements in patients with viral hepatitis with a steady increase of liver stiffness values in parallel with the degree of histological activity [66, 70, 100]. Consistent with these results, the risk of overestimating liver stiffness values has been reported in case of ALT flares in patients with acute viral hepatitis or chronic hepatitis B [101–103], as well as in cases of extrahepatic cholestasis [104] or congestive heart failure [105].

Comparison and combination of approaches

For the diagnosis of significant fibrosis, performances of TE and serum biomarkers have been shown to be equivalent in patients with chronic hepatitis C [56, 69]. For the diagnosis of cirrhosis, the situation is different as TE

appears to be the most accurate non invasive method in HCV patients when compared with currently available biomarkers and routine blood tests [56, 69], allowing the avoidance of liver biopsy in around 90% of cases. TE is thus “the test to be beaten” for those developing alternative methods [106].

In order to increase diagnostic accuracy, new approaches using a combination of TE and serum markers [69, 107, 108] have been proposed. For instance, using an algorithm to evaluate the agreement between TE and FibroTest®, liver biopsy can be avoided for the diagnosis of significant fibrosis in around 75% of patients with chronic hepatitis C [69].

Also, a sequential algorithm (SAFE Biopsy), using APRI as screening test, followed by Fibrotest® in APRI non-classified cases and restricting liver biopsy to patients classified as F0–F1 by non-invasive tests, has been proposed, allowing avoidance of liver biopsy for the diagnosis of significant fibrosis in around 50% of cases [109]. These results have been confirmed in a large-scale, multicenter international validation study of more than 2,000 patients with chronic hepatitis C [110]. Other groups have proposed alternative algorithms combining FibroTest® and APRI [53] either with Forns index (Fibropaca) [111] or Hepascore [112].

In a recent study where these two approaches were compared in the same population of patients with chronic hepatitis C, the number of saved liver biopsies was significantly higher using the combination of TE and Fibrotest® than SAFE Biopsy algorithm for detecting significant fibrosis (72 vs. 48%, respectively; $P < 0.0001$) [113].

A combination of two unrelated methods, such as TE and Fibrotest®, may have certain advantages over the combination of two biomarkers, in that TE provides more direct measurement of the liver structure than serum markers and that there is no relationship between the applicability of TE (success rate and interquartile range) and Fibrotest® (Gilbert’s syndrome, hemolysis, and sepsis) [107].

Novel imaging techniques

New alternative imaging techniques may also become available: magnetic resonance (MR) elastography, which can be implemented readily on standard MR imaging systems with additional hardware; diffusion weighted MR imaging, which measures the apparent diffusion coefficient of water, a parameter that is dependent on the tissue structure; optical digital analysis of computed tomography images of the liver and sonography-based real-time elastography, which can be performed with conventional ultrasound probes during a routine sonography examination

[114]. The theoretical advantages of these methods include the ability to analyze almost the entire liver and the applicability to patients with obesity or ascites. Preliminary studies in human subjects have confirmed the feasibility of these techniques for quantitative assessment of hepatic fibrosis [115–119]. In addition, a recent prospective comparative study in 96 patients with chronic liver disease [120] has suggested that MR elastography had a better diagnostic accuracy than TE for the diagnosis of significant fibrosis (AUROC: 0.99 vs. 0.84, respectively $P < 0.05$). Although such results are encouraging, these techniques remain so far too expensive and time-consuming for implementation in clinical practice for screening hepatic fibrosis.

Very recently, a novel imaging technology (ARFI acoustic radiation force impulse imaging), involving the mechanical excitation of tissue using short-duration ($\sim 262 \mu\text{s}$) acoustic pulses producing shear waves propagation generating localized micron-scale displacements in the tissue, has been proposed [121]. The shear wave velocity (expressed in meters per second) is measured in a region of interest smaller than that of TE (10-mm long and 6-mm wide), but that can be chosen by the examiner. Preliminary results suggest that at least in patients with chronic hepatitis C, its performance is very similar to that of TE, although further validation is warranted [122–126]. The major advantage of this novel technology is that it can be easily implemented on a modified commercial ultrasound machine. However, as compared with LSM values (2.5–75 kPa), ARFI values are in a very narrow range (0.5–4.4 m/s), which could represent a limitation to its use for making decision in clinical practice. More data are also awaited regarding the intra- and inter-observer reproducibility of ARFI.

Conclusion

Significant progress has been made in the non-invasive diagnosis of hepatic fibrosis in patients with chronic hepatitis C. An increasing number of reliable tests are now available: TE, Fibrotest® and APRI have been the most extensively studied and validated. These tests are already widely used in routine clinical practice in France resulting in a significant decrease in the need for liver biopsy [127]. However, it is likely that non-invasive methods will reduce but not completely abolish the need for liver biopsy [28]. The most rational way of using these tools is to make a compromise: use non-invasive methods to classify those patients in whom they perform with high accuracy, limiting liver biopsy to the subset of patients in whom precise non-invasive staging is not possible. In that respect, the use of TE and several patented scores (FibroTest®, Fibrometer®,

and Hepascore[®]) has been recently approved, after an independent systematic review by the French Health Authorities, for the first-line assessment of fibrosis in naïve patients with chronic hepatitis C without co-morbidities [128]. Interpretation of the results of non-invasive methods should always be done critically by an expert clinician according to clinical context and taking into account patient demographics, essential laboratory parameters, and manufacturer's recommendations. Apart from cases where cirrhosis is clinically obvious, a combination of two unrelated methods should be used rather than a single method. A liver biopsy should be performed in case of discordance between results of non-invasive tests, when non-invasive methods are not applicable, or if co-morbidities such as alcoholism or metabolic syndrome are present. In any case, a large specimen (ideally 20–25 mm) should be obtained and reading should be done by an experienced liver pathologist.

In conclusion, we believe that liver biopsy and non-invasive methods, particularly TE, should be employed as an integrated system to allow a more efficient and convenient management of patients with chronic hepatitis C [129].

References

- Ghany MG, Strader DB, Thomas DL, Seeff LB. Diagnosis, management, and treatment of hepatitis C: an update. *Hepatology* 2009;49:1335–1374
- Bravo AA, Sheth SG, Chopra S. Liver biopsy. *N Engl J Med* 2001;344:495–500
- Cadranel JF, Rufat P, Degos F. Practices of liver biopsy in France: results of a prospective nationwide survey. For the Group of Epidemiology of the French Association for the Study of the Liver (AFEF). *Hepatology* 2000;32:477–481
- Castera L, Negre I, Samii K, Buffet C. Pain experienced during percutaneous liver biopsy. *Hepatology* 1999;30:1529–1530
- Castera L, Negre I, Samii K, Buffet C. Patient-administered nitrous oxide/oxygen inhalation provides safe and effective analgesia for percutaneous liver biopsy: a randomized placebo-controlled trial. *Am J Gastroenterol* 2001;96:1553–1557
- Piccino F, Sagnelli E, Pasquale G, Giusti G. Complications following percutaneous liver biopsy. A multicentre retrospective study on 68,276 biopsies. *J Hepatol* 1986;2:165–173
- Bedossa P, Dargère D, Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. *Hepatology* 2003;38:1449–1457
- Regev A, Berho M, Jeffers LJ, Milikowski C, Molina EG, Pyrsopoulos NT, et al. Sampling error and intraobserver variation in liver biopsy in patients with chronic HCV infection. *Am J Gastroenterol* 2002;97:2614–2618
- Rousselet MC, Michalak S, Dupré F, Croué A, Bedossa P, Saint-Andre JP, et al. Sources of variability in histological scoring of chronic viral hepatitis. *Hepatology* 2005;41:257–264
- Bedossa P, Carrat F. Liver biopsy: the best, not the gold standard. *J Hepatol* 2009;50:1–3
- Castera L, Pinzani M. Non-invasive assessment of liver fibrosis: are we ready? *Lancet* 2010;375:1419–1420
- Pinzani M, Vizzutti F, Arena U, Marra F. Technology insight: noninvasive assessment of liver fibrosis by biochemical scores and elastography. *Nat Clin Pract Gastroenterol Hepatol* 2008;5:95–106
- Guechot J, Laudat A, Loria A, Serfaty L, Poupon R, Giboudeau J. Diagnostic accuracy of hyaluronan and type III procollagen amino-terminal peptide serum assays as markers of liver fibrosis in chronic viral hepatitis C evaluated by ROC curve analysis. *Clin Chem* 1996;42:558–563
- McHutchison JG, Blatt LM, de Medina M, Craig JR, Conrad A, Schiff ER, et al. Measurement of serum hyaluronic acid in patients with chronic hepatitis C and its relationship to liver histology. Consensus Interferon Study Group. *J Gastroenterol Hepatol* 2000;15:945–951
- Halfon P, Bourliere M, Penaranda G, Deydier R, Renou C, Botta-Fridlund D, et al. Accuracy of hyaluronic acid level for predicting liver fibrosis stages in patients with hepatitis C virus. *Comp Hepatol* 2005;4:6
- Walsh KM, Fletcher A, MacSween RN, Morris AJ. Basement membrane peptides as markers of liver disease in chronic hepatitis C. *J Hepatol* 2000;32:325–330
- Wong VS, Hughes V, Trull A, Wight DG, Petrik J, Alexander GJ. Serum hyaluronic acid is a useful marker of liver fibrosis in chronic hepatitis C virus infection. *J Viral Hepat* 1998;5:187–192
- Pilette C, Rousselet MC, Bedossa P, Chappard D, Oberti F, Rifflet H, et al. Histopathological evaluation of liver fibrosis: quantitative image analysis vs semi-quantitative scores. Comparison with serum markers. *J Hepatol* 1998;28:439–446
- Oberti F, Valsesia E, Pilette C, Rousselet MC, Bedossa P, Aube C, et al. Noninvasive diagnosis of hepatic fibrosis or cirrhosis. *Gastroenterology* 1997;113:1609–1616
- Saitou Y, Shiraki K, Yamanaka Y, Yamaguchi Y, Kawakita T, Yamamoto N, et al. Noninvasive estimation of liver fibrosis and response to interferon therapy by a serum fibrogenesis marker, YKL-40, in patients with HCV-associated liver disease. *World J Gastroenterol* 2005;11:476–481
- Murawaki Y, Koda M, Okamoto K, Mimura K, Kawasaki H. Diagnostic value of serum type IV collagen test in comparison with platelet count for predicting the fibrotic stage in patients with chronic hepatitis C. *J Gastroenterol Hepatol* 2001;16:777–781
- Murawaki Y, Ikuta Y, Idobe Y, Kawasaki H. Serum matrix metalloproteinase-1 in patients with chronic viral hepatitis. *J Gastroenterol Hepatol* 1999;14:138–145
- Boeker KH, Haberkorn CI, Michels D, Flemming P, Manns MP, Lichtenhagen R. Diagnostic potential of circulating TIMP-1 and MMP-2 as markers of liver fibrosis in patients with chronic hepatitis C. *Clin Chim Acta* 2002;316:71–81
- Pohl A, Behling C, Oliver D, Kilani M, Monson P, Hassanein T. Serum aminotransferase levels and platelet counts as predictors of degree of fibrosis in chronic hepatitis C virus infection. *Am J Gastroenterol* 2001;96:3142–3146
- Imperiale TF, Said AT, Cummings OW, Born LJ. Need for validation of clinical decision aids: use of the AST/ALT ratio in predicting cirrhosis in chronic hepatitis C. *Am J Gastroenterol* 2000;95:2328–2332
- Park GJ, Lin BP, Ngu MC, Jones DB, Katelaris PH. Aspartate aminotransferase: alanine aminotransferase ratio in chronic hepatitis C infection: is it a useful predictor of cirrhosis? *J Gastroenterol Hepatol* 2000;15:386–390
- Sheth SG, Flamm SL, Gordon FD, Chopra S. AST/ALT ratio predicts cirrhosis in patients with chronic hepatitis C virus infection. *Am J Gastroenterol* 1998;93:44–48
- Sebastiani G, Alberti A. Non invasive fibrosis biomarkers reduce but not substitute the need for liver biopsy. *World J Gastroenterol* 2006;12:3682–3694

29. Castera L. Assessing liver fibrosis. *Expert Rev Gastroenterol Hepatol* 2008;2:541–552.
30. Manning DS, Afdhal NH. Diagnosis and quantitation of fibrosis. *Gastroenterology* 2008;134:1670–1681.
31. Mehta P, Ploutz-Snyder R, Nandi J, Rawlins SR, Sanderson SO, Levine RA. Diagnostic accuracy of serum hyaluronic acid, FIBROspect II, and YKL-40 for discriminating fibrosis stages in chronic hepatitis C. *Am J Gastroenterol* 2008;103:928–936.
32. Lackner C, Struber G, Liegl B, Leibl S, Ofner P, Bankuti C, et al. Comparison and validation of simple noninvasive tests for prediction of fibrosis in chronic hepatitis C. *Hepatology* 2005;41:1376–1382.
33. Imbert-Bismut F, Ratiu V, Pieroni L, Charlotte F, Benhamou Y, Poynard T. Biochemical markers of liver fibrosis in patients with hepatitis C virus infection: a prospective study. *Lancet* 2001;357:1069–1075.
34. Forns X, Ampurdanes S, Llovet JM, Aponte J, Quinto L, Martinez-Bauer E, et al. Identification of chronic hepatitis C patients without hepatic fibrosis by a simple predictive model. *Hepatology* 2002;36:986–992.
35. Wai CT, Greenson JK, Fontana RJ, Kalbfleisch JD, Marrero JA, Conjeevaram HS, et al. A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. *Hepatology* 2003;38:518–526.
36. Patel K, Gordon SC, Jacobson I, Hezode C, Oh E, Smith KM, et al. Evaluation of a panel of non-invasive serum markers to differentiate mild from moderate-to-advanced liver fibrosis in chronic hepatitis C patients. *J Hepatol* 2004;41:935–942.
37. Leroy V, Monier F, Bottari S, Trocme C, Sturm N, Hilleret MN, et al. Circulating matrix metalloproteinases 1, 2, 9 and their inhibitors TIMP-1 and TIMP-2 as serum markers of liver fibrosis in patients with chronic hepatitis C: comparison with PIIINP and hyaluronic acid. *Am J Gastroenterol* 2004;99:271–279.
38. Rosenberg WM, Voelker M, Thiel R, Becka M, Burt A, Schuppan D, et al. Serum markers detect the presence of liver fibrosis: a cohort study. *Gastroenterology* 2004;127:1704–1713.
39. Sud A, Hui JM, Farrell GC, Bandara P, Kench JG, Fung C, et al. Improved prediction of fibrosis in chronic hepatitis C using measures of insulin resistance in a probability index. *Hepatology* 2004;39:1239–1247.
40. Adams LA, Bulsara M, Rossi E, DeBoer B, Speers D, George J, et al. Hepascore: an accurate validated predictor of liver fibrosis in chronic hepatitis C infection. *Clin Chem* 2005;51:1867–1873.
41. Cales P, Oberti F, Michalak S, Hubert-Fouchard I, Rousselet MC, Konate A, et al. A novel panel of blood markers to assess the degree of liver fibrosis. *Hepatology* 2005;42:1373–1381.
42. Fontana RJ, Kleiner DE, Bilionick R, Terrault N, Afdhal N, Belle SH, et al. Modeling hepatic fibrosis in African American and Caucasian American patients with chronic hepatitis C virus infection. *Hepatology* 2006;44:925–935.
43. Koda M, Matunaga Y, Kawakami M, Kishimoto Y, Suou T, Murawaki Y. FibroIndex, a practical index for predicting significant fibrosis in patients with chronic hepatitis C. *Hepatology* 2007;45:297–306.
44. Callewaert N, Van Vlierberghe H, Van Hecke A, Laroy W, Delanghe J, Contreras R. Noninvasive diagnosis of liver cirrhosis using DNA sequencer-based total serum protein glycemics. *Nat Med* 2004;10:429–434.
45. Lok AS, Ghany MG, Goodman ZD, Wright EC, Everson GT, Sterling RK, et al. Predicting cirrhosis in patients with hepatitis C based on standard laboratory tests: results of the HALT-C cohort. *Hepatology* 2005;42:282–292.
46. Islam S, Antonsson L, Westin J, Lagging M. Cirrhosis in hepatitis C virus-infected patients can be excluded using an index of standard biochemical serum markers. *Scand J Gastroenterol* 2005;40:867–872.
47. Vallet-Pichard A, Mallet V, Nalpas B, Verkarre V, Nalpas A, Dhalluin-Venier V, et al. FIB-4: an inexpensive and accurate marker of fibrosis in HCV infection. Comparison with liver biopsy and fibrotest. *Hepatology* 2007;46:32–36.
48. Fontana RJ, Goodman ZD, Dienstag JL, Bonkovsky HL, Nai-shadham D, Sterling RK, et al. Relationship of serum fibrosis markers with liver fibrosis stage and collagen content in patients with advanced chronic hepatitis C. *Hepatology* 2008;47:789–798.
49. Poynard T, Morra R, Halfon P, Castera L, Ratiu V, Imbert Bismut F, et al. Meta-analyses of Fibrotest diagnostic value in chronic liver disease. *BMC Gastroenterol* 2007;7:40.
50. Shaheen AA, Myers RP. Diagnostic accuracy of the aspartate aminotransferase-to-platelet ratio index for the prediction of hepatitis C-related fibrosis: a systematic review. *Hepatology* 2007;46:912–921.
51. Parkes J, Guha IN, Roderick P, Rosenberg W. Performance of serum marker panels for liver fibrosis in chronic hepatitis C. *J Hepatol* 2006;44:462–474.
52. Halfon P, Bacq Y, De Muret A, Penaranda G, Bourliere M, Ouzan D, et al. Comparison of test performance profile for blood tests of liver fibrosis in chronic hepatitis C. *J Hepatol* 2007;46:395–402.
53. Leroy V, Hilleret MN, Sturm N, Trocme C, Renversez JC, Faure P, et al. Prospective comparison of six non-invasive scores for the diagnosis of liver fibrosis in chronic hepatitis C. *J Hepatol* 2007;46:775–782.
54. Leroy V, Halfon P, Bacq Y, Boursier J, Rousselet MC, Bourliere M, et al. Diagnostic accuracy, reproducibility and robustness of fibrosis blood tests in chronic hepatitis C: a meta-analysis with individual data. *Clin Biochem* 2008;41:1368–1376.
55. Cales P, de Ledinghen V, Halfon P, Bacq Y, Leroy V, Boursier J, et al. Evaluating the accuracy and increasing the reliable diagnosis rate of blood tests for liver fibrosis in chronic hepatitis C. *C. Liver Int* 2008;28:1352–1362.
56. Degos F, Perez P, Roche B, Mahmoudi A, Asselineau J, Voitot H, et al. Diagnostic accuracy of FibroScan and comparison to liver fibrosis biomarkers in chronic viral hepatitis: a multicenter prospective study (the FIBROSTIC study). *J Hepatol* 2010;53:1013–1021.
57. Fraser JR, Gibson PR. Mechanisms by which food intake elevates circulating levels of hyaluronan in humans. *J Intern Med* 2005;258:460–466.
58. Piton A, Poynard T, Imbert-Bismut F, Khalil L, Delattre J, Pelissier E, et al. Factors associated with serum alanine transaminase activity in healthy subjects: consequences for the definition of normal values, for selection of blood donors, and for patients with chronic hepatitis C. MULTIVIRC Group. *Hepatology* 1998;27:1213–1219.
59. Cales P, Veillon P, Konate A, Mathieu E, Ternisien C, Chevailier A, et al. Reproducibility of blood tests of liver fibrosis in clinical practice. *Clin Biochem* 2008;41:10–18.
60. Halfon P, Imbert-Bismut F, Messous D, Antoniotti G, Benchettit D, Cart-Lamy P, et al. A prospective assessment of the inter-laboratory variability of biochemical markers of fibrosis (FibroTest) and activity (ActiTest) in patients with chronic liver disease. *Comp Hepatol* 2002;1:3.
61. Imbert-Bismut F, Messous D, Thibaut V, Myers RB, Piton A, Thabut D, et al. Intra-laboratory analytical variability of biochemical markers of fibrosis (Fibrotest) and activity (Actitest) and reference ranges in healthy blood donors. *Clin Chem Lab Med* 2004;42:323–333.
62. Poynard T, Munteanu M, Imbert-Bismut F, Charlotte F, Thabut D, Le Calvez S, et al. Prospective analysis of discordant results between biochemical markers and biopsy in patients with chronic hepatitis C. *Clin Chem* 2004;10:10.

63. Sandrin L, Fourquet B, Hasquenoph JM, Yon S, Fournier C, Mal F, et al. Transient elastography: a new noninvasive method for assessment of hepatic fibrosis. *Ultrasound Med Biol* 2003;29:1705–1713.
64. Castera L, Forns X, Alberti A. Non-invasive evaluation of liver fibrosis using transient elastography. *J Hepatol* 2008;48:835–847.
65. Roulot D, Czernichow S, Le Clesiau H, Costes JL, Vergnaud AC, Beaugrand M. Liver stiffness values in apparently healthy subjects: influence of gender and metabolic syndrome. *J Hepatol* 2008;48:606–613.
66. Fraquelli M, Rigamonti C, Casazza G, Conte D, Donato MF, Ronchi G, et al. Reproducibility of transient elastography in the evaluation of liver fibrosis in patients with chronic liver disease. *Gut* 2007;56:968–973.
67. Boursier J, Konate A, Guilluy M, Gorea G, Sawadogo A, Quemener E, et al. Learning curve and interobserver reproducibility evaluation of liver stiffness measurement by transient elastography. *Eur J Gastroenterol Hepatol* 2008;20:693–701.
68. Ziolkowski M, Handra-Luka A, Kettaneh A, Christidis C, Mal F, Kazemi F, et al. Noninvasive assessment of liver fibrosis by measurement of stiffness in patients with chronic hepatitis C. *Hepatology* 2005;41:48–54.
69. Castera L, Vergniol J, Foucher J, Le Bail B, Chanteloup E, Haaser M, et al. Prospective comparison of transient elastography, Fibrotest, APRI, and liver biopsy for the assessment of fibrosis in chronic hepatitis C. *Gastroenterology* 2005;128:343–350.
70. Arena U, Vizzutti F, Abraldes JG, Corti G, Stasi C, Moscarella S, et al. Reliability of transient elastography for the diagnosis of advanced fibrosis in chronic hepatitis C. *Gut* 2008;57:1288–1293.
71. Lupsor M, Badea R, Stefanescu H, Grigorescu M, Sparchez Z, Serban A, et al. Analysis of histopathological changes that influence liver stiffness in chronic hepatitis C. Results from a cohort of 324 patients. *J Gastrointest Liver Dis* 2008;17:155–163.
72. Sporea I, Sirli R, Deleanu A, Tudora A, Curescu M, Cornianu M, et al. Comparison of the liver stiffness measurement by transient elastography with the liver biopsy. *World J Gastroenterol* 2008;14:6513–6517.
73. Shaheen AA, Wan AF, Myers RP. FibroTest and FibroScan for the prediction of hepatitis C-related fibrosis: a systematic review of diagnostic test accuracy. *Am J Gastroenterol* 2007;102:2589–2600.
74. Talwalkar JA, Kurtz DM, Schoenleber SJ, West CP, Montori VM. Ultrasound-based transient elastography for the detection of hepatic fibrosis: systematic review and meta-analysis. *Clin Gastroenterol Hepatol* 2007;5:1214–1220.
75. Friedrich-Rust M, Ong MF, Martens S, Sarrazin C, Bojunga J, Zeuzem S, et al. Performance of transient elastography for the staging of liver fibrosis: a meta-analysis. *Gastroenterology* 2008;134:960–974.
76. Ganne-Carrie N, Ziolkowski M, de Ledinghen V, Douvin C, Marcellin P, Castera L, et al. Accuracy of liver stiffness measurement for the diagnosis of cirrhosis in patients with chronic liver diseases. *Hepatology* 2006;44:1511–1517.
77. Foucher J, Chanteloup E, Vergniol J, Castera L, Le Bail B, Adhoute X, et al. Diagnosis of cirrhosis by transient elastography (FibroScan): a prospective study. *Gut* 2006;55:403–408.
78. Carrion JA, Navasa M, Bosch J, Bruguera M, Gilabert R, Forns X. Transient elastography for diagnosis of advanced fibrosis and portal hypertension in patients with hepatitis C recurrence after liver transplantation. *Liver Transplant* 2006;12:1791–1798.
79. Vizzutti F, Arena U, Romanelli RG, Rega L, Foschi M, Colagrande S, et al. Liver stiffness measurement predicts severe portal hypertension in patients with HCV-related cirrhosis. *Hepatology* 2007;45:1290–1297.
80. Ripoll C, Groszmann R, Garcia-Tsao G, Grace N, Burroughs A, Planas R, et al. Hepatic venous pressure gradient predicts clinical decompensation in patients with compensated cirrhosis. *Gastroenterology* 2007;133:481–488.
81. Kazemi F, Kettaneh A, N'Kontchou G, Pinto E, Ganne-Carrie N, Trinchet JC, et al. Liver stiffness measurement selects patients with cirrhosis at risk of bearing large oesophageal varices. *J Hepatol* 2006;45:230–235.
82. Bureau C, Metivier S, Peron JM, Selves J, Robic MA, Gourraud PA, et al. Transient elastography accurately predicts presence of significant portal hypertension in patients with chronic liver disease. *Aliment Pharmacol Ther* 2008;27:1261–1268.
83. Castera L, Le Bail B, Roudot-Thoraval F, Bernard PH, Foucher J, Merrouche W, et al. Early detection in routine clinical practice of cirrhosis and oesophageal varices in chronic hepatitis C: comparison of transient elastography (FibroScan) with standard laboratory tests and non-invasive scores. *J Hepatol* 2009;50:59–68.
84. Pineda JA, Recio E, Camacho A, Macias J, Almodovar C, Gonzalez-Serrano M, et al. Liver stiffness as a predictor of esophageal varices requiring therapy in HIV/hepatitis C virus-coinfected patients with cirrhosis. *J Acquir Immune Defic Syndr* 2009;51:445–449.
85. Castera L. Elastography in the non-invasive evaluation of the extent of fibrosis and in the diagnosis of portal hypertension. In Franchis RD, editor. Portal Hypertension V: Proceedings of the Fifth Baveno International Consensus Workshop. Wiley-Blackwell; 2010. p. 18–27.
86. Rigamonti C, Donato MF, Fraquelli M, Agnelli F, Ronchi G, Casazza G, et al. Transient elastography predicts fibrosis progression in patients with recurrent hepatitis C after liver transplantation. *Gut* 2008;57:821–827.
87. Corradi F, Piscaglia F, Flori S, D'Errico-Grigioni A, Vasuri F, Tame MR, et al. Assessment of liver fibrosis in transplant recipients with recurrent HCV infection: usefulness of transient elastography. *Dig Liver Dis* 2009;41:217–225.
88. Harada N, Soejima Y, Taketomi A, Yoshizumi T, Ikegami T, Yamashita Y, et al. Assessment of graft fibrosis by transient elastography in patients with recurrent hepatitis C after living donor liver transplantation. *Transplantation* 2008;85:69–74.
89. Beckebaum S, Iacob S, Klein CG, Dechene A, Varghese J, Baba HA, et al. Assessment of allograft fibrosis by transient elastography and noninvasive biomarker scoring systems in liver transplant patients. *Transplantation* 2010;89:983–993.
90. Carrion JA, Torres F, Crespo G, Miquel R, Garcia-Valdecasas JC, Navasa M, et al. Liver stiffness identifies two different patterns of fibrosis progression in patients with hepatitis C virus recurrence after liver transplantation. *Hepatology* 2010;51:23–34.
91. Cholongitas E, Tsochatzis E, Goulis J, Burroughs AK. Noninvasive tests for evaluation of fibrosis in HCV recurrence after liver transplantation: a systematic review. *Transpl Int* 2010;23:861–870.
92. Hézode C, Castéra L, Roudot-Thoraval F, Rosa I, Roulot D, Leroy V, et al. Prospective evaluation of liver stiffness dynamics during and after peginterferon alpha-ribavirin treatment in patients with chronic hepatitis C (abstract). *J Hepatol* 2009;50 Suppl 1:S226.
93. Ogawa E, Furusyo N, Toyoda K, Takeoka H, Maeda S, Hayashi J. The longitudinal quantitative assessment by transient elastography of chronic hepatitis C patients treated with pegylated interferon alpha-2b and ribavirin. *Antiviral Res* 2009;83:127–134.

94. Vergniol J, Foucher J, Castera L, Bernard PH, Tournan R, Terrebonne E, et al. Changes of non-invasive markers and FibroScan values during HCV treatment. *J Viral Hepat* 2009;16:132–140.
95. Masuzaki R, Tateishi R, Yoshida H, Goto E, Sato T, Ohki T, et al. Prospective risk assessment for hepatocellular carcinoma development in patients with chronic hepatitis C by transient elastography. *Hepatology* 2009;49:1954–1961.
96. Castera L. Liver stiffness and hepatocellular carcinoma: liaisons danger uses? *Hepatology* 2009;49:1793–1794.
97. Castera L, Foucher J, Bernard PH, Carvalho F, Allaix D, Merrouche W, et al. Pitfalls of liver stiffness measurement: a 5-year prospective study of 13,369 examinations. *Hepatology* 2010;51:828–835.
98. Kettaneh A, Marcellin P, Douvin C, Poupon R, Zioli M, Beaugrand M, et al. Features associated with success rate and performance of FibroScan measurements for the diagnosis of cirrhosis in HCV patients: a prospective study of 935 patients. *J Hepatol* 2007;46:628–634.
99. Lucidarme D, Foucher J, Le Bail B, Vergniol J, Castera L, Duburque C, et al. Factors of accuracy of transient elastography (fibroscan) for the diagnosis of liver fibrosis in chronic hepatitis C. *Hepatology* 2009;1083–1089.
100. Chan HL, Wong GL, Choi PC, Chan AW, Chim AM, Yiu KK, et al. Alanine aminotransferase-based algorithms of liver stiffness measurement by transient elastography (Fibroscan) for liver fibrosis in chronic hepatitis B. *J Viral Hepat* 2009;16:36–44.
101. Coco B, Oliveri F, Maina AM, Ciccorossi P, Sacco R, Colombo P, et al. Transient elastography: a new surrogate marker of liver fibrosis influenced by major changes of transaminases. *J Viral Hepat* 2007;14:360–369.
102. Sagir A, Erhardt A, Schmitt M, Haussinger D. Transient elastography is unreliable for detection of cirrhosis in patients with acute liver damage. *Hepatology* 2007;47:592–595.
103. Arena U, Vizzutti F, Corti G, Ambu S, Stasi C, Bresci S, et al. Acute viral hepatitis increases liver stiffness values measured by transient elastography. *Hepatology* 2008;47:380–384.
104. Millonig G, Reimann FM, Friedrich S, Fonouni H, Mehrabi A, Buchler MW, et al. Extrahepatic cholestasis increases liver stiffness (FibroScan) irrespective of fibrosis. *Hepatology* 2008;48:1718–1723.
105. Millonig G, Friedrich S, Adolf S, Fonouni H, Golriz M, Mehrabi A, et al. Liver stiffness is directly influenced by central venous pressure. *J Hepatol* 2010;52:206–210.
106. Bosch J. Towards the non-invasive diagnosis of cirrhosis: the nuts-cirrhosis connection. *J Hepatol* 2009;50:4–6.
107. Poynard T, Ingiliz P, Elkrief L, Munteanu M, Lebray P, Morra R, et al. Concordance in a world without a gold standard: a new non-invasive methodology for improving accuracy of fibrosis markers. *PLoS ONE* 2008;3:e3857.
108. Boursier J, Vergniol J, Sawadogo A, Dakka T, Michalak S, Gallois Y, et al. The combination of a blood test and Fibroscan improves the non-invasive diagnosis of liver fibrosis. *Liver Int* 2009;29:1507–1515.
109. Sebastiani G, Vario A, Guido M, Noventa F, Plebani M, Pistis R, et al. Stepwise combination algorithms of non-invasive markers to diagnose significant fibrosis in chronic hepatitis C. *J Hepatol* 2006;44:686–693.
110. Sebastiani G, Halfon P, Castera L, Pol S, Thomas DL, Mangia A, et al. SAFE biopsy: a validated method for large-scale staging of liver fibrosis in chronic hepatitis C. *Hepatology* 2009;49:1821–1827.
111. Bourliere M, Penaranda G, Renou C, Botta-Fridlund D, Tran A, Portal I, et al. Validation and comparison of indexes for fibrosis and cirrhosis prediction in chronic hepatitis C patients: proposal for a pragmatic approach classification without liver biopsies. *J Viral Hepat* 2006;13:659–670.
112. Bourliere M, Penaranda G, Ouzan D, Renou C, Botta-Fridlund D, Tran A, et al. Optimized stepwise combination algorithms of non-invasive liver fibrosis scores including Hepascore in hepatitis C virus patients. *Aliment Pharmacol Ther* 2008;28:458–467.
113. Castera L, Sebastiani G, Le Bail B, de Ledinghen V, Couzigou P, Alberti A. Prospective comparison of two algorithms combining non-invasive methods for staging liver fibrosis in chronic hepatitis C. *J Hepatol* 2010;52:191–198.
114. Talwalkar JA, Yin M, Fidler JL, Sanderson SO, Kamath PS, Ehman RL. Magnetic resonance imaging of hepatic fibrosis: emerging clinical applications. *Hepatology* 2008;47:332–342.
115. Huwart L, Sempoux C, Salameh N, Jamart J, Annet L, Sinkus R, et al. Liver fibrosis: noninvasive assessment with MR elastography versus aspartate aminotransferase-to-platelet ratio index. *Radiology* 2007;245:458–466.
116. Yin M, Talwalkar JA, Glaser KJ, Manduca A, Grimm RC, Rossman PJ, et al. Assessment of hepatic fibrosis with magnetic resonance elastography. *Clin Gastroenterol Hepatol* 2007;5:1207–1213.
117. Friedrich-Rust M, Ong MF, Herrmann E, Dries V, Samaras P, Zeuzem S, et al. Real-time elastography for noninvasive assessment of liver fibrosis in chronic viral hepatitis. *AJR Am J Roentgenol* 2007;188:758–764.
118. Lewin M, Poujol-Robert A, Boelle PY, Wendum D, Lasnier E, Viallon M, et al. Diffusion-weighted magnetic resonance imaging for the assessment of fibrosis in chronic hepatitis C. *Hepatology* 2007;46:658–665.
119. Romero-Gomez M, Gomez-Gonzalez E, Madrazo A, Vera-Valencia M, Rodrigo L, Perez-Alvarez R, et al. Optical analysis of computed tomography images of the liver predicts fibrosis stage and distribution in chronic hepatitis C. *Hepatology* 2008;47:810–816.
120. Huwart L, Sempoux C, Vicaut E, Salameh N, Annet L, Danse E, et al. Magnetic resonance elastography for the noninvasive staging of liver fibrosis. *Gastroenterology* 2008;135:32–40.
121. Castera L. Acoustic radiation force impulse imaging: a new technology for the noninvasive assessment of liver fibrosis? *J Gastrointest Liver Dis* 2009;18:411–412.
122. Friedrich-Rust M, Wunder K, Kriener S, Sotoudeh F, Richter S, Bojunga J, et al. Liver fibrosis in viral hepatitis: noninvasive assessment with acoustic radiation force impulse imaging versus transient elastography. *Radiology* 2009;252:595–604.
123. Lupsor M, Badea R, Stefanescu H, Sparchez Z, Branda H, Serban A, et al. Performance of a new elastographic method (ARFI technology) compared to unidimensional transient elastography in the noninvasive assessment of chronic hepatitis C. Preliminary results. *J Gastrointest Liver Dis* 2009;18:303–310.
124. Fierbinteanu-Braticevici C, Andronescu D, Usvat R, Cretoiu D, Baicus C, Marinatoschi G. Acoustic radiation force imaging sonelastography for noninvasive staging of liver fibrosis. *World J Gastroenterol* 2009;15:5525–5532.
125. Boursier J, Isselin G, Fouchard-Hubert I, Oberti F, Dib N, Lebigot J, et al. Acoustic radiation force impulse: a new ultrasonographic technology for the widespread noninvasive diagnosis of liver fibrosis. *Eur J Gastroenterol Hepatol* 2010;22:1074–1084.
126. Takahashi H, Ono N, Eguchi Y, Eguchi T, Kitajima Y, Kawaguchi Y, et al. Evaluation of acoustic radiation force impulse elastography for fibrosis staging of chronic liver disease: a pilot study. *Liver Int* 2010;30:538–545.
127. Castera L, Denis J, Babany G, Roudot-Thoraval F. Evolving practices of non-invasive markers of liver fibrosis in patients with chronic hepatitis C in France: time for new guidelines? *J Hepatol* 2007;46:528–529.
128. Non invasive methods for the evaluation of hepatic fibrosis/cirrhosis: an update. www.has-sante.fr (2008).
129. Castera L, Pinzani M. Biopsy and non-invasive methods for the diagnosis of liver fibrosis: does it take two to tango? *Gut* 2010;59:861–866.